A NEW TRIOXYGENATED DITERPENE FROM THE MOLLUSK APLYSIA DACTYLOMELA

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Diterpenes with a dolabellane skeleton were initially isolated from the herbivorous sea hare *Dolabella californica* (1). They were later isolated from the brown algae of the family Dyctiotaceae (2-4), from the sea whip *Eunicea caliculata* (5), and from the mollusk *Aplysia dactylomela* Rang (Gasteropod, Opisthobranch) (6). From a new collection of *A. dactylomela*, a new diterpene having the dolabellane skeleton was isolated.

RESULTS AND DISCUSSION

Compound 1 $[\alpha]D^{25}+13^{\circ}$ (c 0.06, CHCl₃), had the molecular formula $C_{24}H_{38}O_5$ from the mass measurement of the parent ion, M^+ , at m/z 406. The presence of two methyl signals at δ 1.9 and 2.0, a band at 1730 cm^{-1} in the ir spectrum, and signals in the ¹³C-nmr spectrum at 69.9 (d) and 84.8 (s) ppm strongly suggested that 1 was a diacetate. From the rest of the spectroscopic data of this compound (see Experimental section), we inferred that it is bicyclic with the skeleton of dolabellane. The structure was chemically confirmed by base catalyzed hydrolysis to the previously isolated diol 2, whose structure was rigorously determined by X-ray diffraction analysis (6). The stereochemis-



try of **1** was deduced from the following evidence. The NOESY spectrum exhibited the presence of nOes indicating that the bridgehead Me-15, H-3, and H-10 have a β orientation, while the Me-16, Me-17, H-2, and H-11 are α (Figure 1).



EXPERIMENTAL

COLLECTION, EXTRACTION, AND CHRO-MATOGRAPHIC SEPARATION .--- For general experimental procedures, see González et al. (7). In all, 35 specimens of A. dactylomela were collected in August 1983, at Los Cristianos, Tenerife, Canary Islands. A specimen of the animal was deposited at the Department of Zoology, University of La Laguna. The hepatopancreas of each animal was removed and stored in Me₂CO for 2 days at 10°. The solvent was evaporated to leave a darkgreen viscous oil (40 g). The crude extract was chromatographed on Si gel using a solvent gradient of increasing polarity from n-hexane to EtOAc. The fraction eluted with n-hexane-EtOAc (9:1) was rechromatographed with low pressure Si gel chromatography using n-hexane as eluent to obtain the diacetate 1 (20 mg) as an oil, $[\alpha]D^{25} + 13^{\circ}$ (c 0.06, CHCl₃); ir (CHCl₃) 3000, 2960, 2920, 2850, 1730, 1450, 1250 cm⁻¹; ¹H nmr (200 MHz, δ, CDCl₃) 0.80 (s, Me-15), 0.85 (m, H-14); 1.02 (d, Me-16, J=6.5 Hz), 1.23 (m, H-9β), 1.37 (s, Me-20), 1.40 (s, Me-17), 1.5 (m, H-6), 1.5 (m, H-13), 1.63 (s, Me-19), 1.80 (m, H-11), 1.90 (m, H-5), 1.90 [s, Me(OAc)], 2.00 [s, Me(OAc)], 2.47 (dd, H-9a, J=11.5 and 13 Hz), 2.5 (m, H-4), 3.07 (d, H-7, J=9 Hz), 3.18 (ddd, H-12, J=11.5 and 5 Hz), 4.97 (bd, H-10, J=9.3 Hz), 5.21 (dd, H-2,

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J=16.3 and 2 Hz), 5.69 (dd, H-3, J=16.3 and 4.5 Hz); 13 C-nmr (CDCl₃) 15.9 (q), 16.8 (q), 20.7 (q), 21.1 (q), 22.8 (t), 23.2 (q), 23.4 (q), 32.3 (t), 33.5 (d), 39.9 (t), 44.3 (d), 44.5 (t), 48.9 (s), 50.6 (d), 60.4 (s), 63.4 (d), 69.9 (d), 84.8 (s), 134.2 (d), 136 (d), 170.3 (s); ms M⁺ at m/z 406 (4), 346 (8), 286 (53), 271 (100), 243 (56), 228 (25).

HYDROLYSIS OF DIACETATE 1.—A mixture of the diacetate 1 (15 mg, 0.037 mmol) and K_2CO_3 (15 mg) in MeOH (2 ml) was allowed to stand at room temperature for 24 h. The mixture was neutralized with HCl 1 N and extracted with Et_2O (3×10 ml). The Et_2O extracts were combined and dried over anhydrous MgSO₄. The solvent was evaporated to yield 10 mg of the diol 2, identical (mp, tlc, ¹H-nmr) with authentic material.

ACKNOWLEDGMENTS

This research was supported by Grant 3481/83 by the Spanish CAICYT. José J. Fernández thanks the Spanish Ministry of Education and Science for F.P.I. fellowship. Fernando Cataldo thanks AIETI Foundation, Madrid, for a fellowship.

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Received 1 May 1987